

Review Article: Ovarian Cancer

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ARTICLE INFO	ABSTRACT
Received: March 15, 2024 Accepted: April 22, 2024 Volume: 4 Issue: 1 KEYWORDS Ovary, Gynecological malignancy, Mutation, Molecular genetics, Signaling pathway	The first record of cancer dates back to Ancient Egypt in 3000 BC, and it has been studied throughout history as an ancient illness. Oncology is the study of cancer, and during the past few decades, significant progress has been made in this area. Despite these initiatives, cancer still claims the lives of almost 10 million people annually. Since cancer is caused by a series of faults in one's own cells, finding a cure for it involves figuring out how to get rid of the malignant cells without harming one's own body. The diagnosis and treatment of ovarian cancer are the most challenging of all gynecological cancers. Due to its nearly symptomless early phases, this type of gynecological cancer is either discovered by chance for example, during a diagnostic laparoscopy or is detected at an advanced stage when symptoms start to manifest. Our capacity to detect and successfully treat this illness at a period when it is still curable is restricted by an insufficient understanding of early molecular events in ovarian carcinogenesis. Due to the aforementioned factors, ovarian cancer has acquired the moniker in the United States of America (The Silent Killer). This phenomenon results from the exceedingly limited early diagnostic resources (biomarkers) that we currently have. Given its high mortality rate, gynecological oncology research is heavily focused in this area. To promote growth, cells with mutant genotypes are chosen and then gradually altered, which might result in the growth of tumors. Spontaneous mutations are uncommon during each cell generation because to the rigorous mechanism for the detection of DNA errors and repair, and multiple mutations are required to orchestrate tumor formation. Once it has begun, the mutational accumulation is sped up by increased susceptibility to mutagenic agents, a breakdown of some aspect of the mutagenic repair system, or a combination of both. The Wnt/β-catenin canonical signaling pathway plays a role in cancer, wound healing, and developmental processes. However, research on the role of Wnt/canonical signaling in the development of ovarian cancer has just lately started to surface. Hence, the present focused on the most genes participated in ovarian cancer and the signaling pathway of the most important gene, Wnt/β-catenin.

1. Normal Ovarian Physiology

On each side of the pelvis, there are two paired organs called the ovaries, which each one weighs about 4-8 grams and has dimensions of 2.5–5 cm in length and 1.5–3 cm in diameter. The ovaries have two medial and lateral surfaces, two anterior or mesovarium borders, two posterior or free borders, and two tubal and uterine poles. The epithelial coat of the ovaries and the posterosuperior layer of the broad ligament are both continuous with the two layers of peritoneum that make up the mesovarium (1). The ovarian arteries provide the ovaries with blood, and hypothalamus controls the secretion of gonadotropin-releasing hormone (GnRH) that released by females in a cyclical fashion to preserve the menstrual cycle (2). To promote ovulation and preserve the corpus luteum, luteinizing hormone (LH) acts on ovarian follicle that develops as a result of follicle stimulating hormone (FSH) that encourages the release of progesterone and estradiol. Through a negative feedback loop, these steroidal hormones can prevent releasing of GnRH and prevents releasing of FSH and LH. However, estradiol induces an LH surge that is connected to ovulation when it is present in sustained high levels (3). Estradiol and testosterone's bioavailability is altered by the impacts of leptin, a hormone produced by fat cells, and insulin, a hormone produced by the pancreas, through changes in the liver's production of sex hormone-binding globulin (SHBG). The ovary can also be directly affected by insulin (4, 5).

2. Ovarian Cancer

2.1. Introduction

Ovarian cancer accounts for 6% of all female cancer deaths in the United States, making it the sixth most common cancer-related cause of death for females (6). Although, advancements in treatment of the disease continues to be the most fatal gynecological malignancy, approximately 69% of overall patients pass away from illness, in comparison with 19% of patients. It is mostly attributed for delaying detection brought on by a lack of an efficient screening plan; however, only 20% of individual patients have identified when tumor remain restricted to disease (7, 8). Hence, traditional surgery and chemotherapy can cure more than 80% of patients. Age, obesity, ovarian cancer in the family or other cancers act as risk factors for disease (9). Current accepted theory holds that ovulation repeatedly damages DNA and then repairs it, causing irreversible oxidative DNA damage that either triggers or facilitates the development of ovarian surface epithelial cells (OSE) tumors (10). Serous, mucinous, endometrioid, and clear cell histological subtypes of ovarian neoplasms are subdivided (11). The BRCA1, BRCA2, MLH1 and MSH2 genes are linked to hereditary frequent germline mutations. Development, wound healing, and cancer are all impacted by the Wnt/ β -catenin canonical signaling pathway (12). It's interesting to note that all of these processes seem to be connected, either directly or indirectly, to the development of ovarian cancer. It has been shown that different ovaries have distinctive Wnt signaling molecule expression patterns, and that certain Wnts are expressed in human ovarian cancer cell lines. Further evidence that different ovarian cancer subtypes exhibit unique genetic abnormalities of Wnt/canonical signaling pathway components comes from reports of variations in the expression and function of Wnt/canonical signaling pathway components in normal vs malignant OSE cells (13, 14). Different studies predicted that ovarian cancer cells have abnormalities in Wnt/canonical signaling, which may cause and/or impact ovarian tumorigenesis. Also, it showed that Wnts can secrete glycoproteins that interact with Frizzled (Fz) seven-transmembrane spanning receptors to either cause the cytosol to accumulate β -catenin and the subsequent co-activation of target genes during transcription, to release intracellular Ca^{2+} , or to activate protein kinases (15-17). The canonical pathway involves the binding of Wnts to Fz receptors along with the co-receptors LDL-receptor related protein (15). This inhibits the β -catenin phosphorylation by the serine / threonine kinase, glycogen synthase kinase 3 (GSK3) within a large cytoplasmic complex (18). Inhibiting β -catenin phosphorylation prevents it from being degraded by the ubiquitin/proteasome system, which leads to a buildup of the cytosolic molecule in its uncomplexed state. Simple β -catenin then moves to the nucleus where it interacts with T-cell factor/lymphoid enhancer factor (TCF/LEF) to activate target genes like c-myc, l-myc, cyclin D, vascular endothelial growth factor (VEGF), bone morphogenetic protein 4 (BMP4), E-cadherin, and N-cadherin (19, 20). Aberrations in the Wnt/canonical pathway cause certain target genes to be inappropriately activated or silenced by β -catenin, which permits unchecked cell proliferation and/or changes in cell differentiation. β -catenin molecules can be involved in Wnt/canonical pathway as well as being sequestered to plasma membrane as a result of their interactions with cadherin cell adhesion molecules and β -catenin (21). Therefore, it has been suggested that the β -catenin molecules possess particular cadherin and β -catenin binding sites, even though the mechanism behind the selection of β -catenin molecules to the plasma membrane vs the cell nucleus has not yet been fully understood (22).

The expression of Wnt ligand and Fz receptor has been proven in earlier investigations. In addition, different reports demonstrated that ovarian cancers might precipitate β -catenin and LEF-1 (23, 24). These data imply that although the canonical signaling pathway may be more activated in ovarian cancer cells, components of the Wnt/canonical signaling pathway are present in both healthy and malignant OSE. Approximately, 85% of mucinous carcinomas have ovarian serous carcinomas according to analyses of genetic mutations according to ovarian cancer histological subtypes (25). About 40% of ovarian endometrioid adenocarcinomas (OBAs) have mutations in the *CTNNB1* gene, which codes for β -catenin, which affect Wnt signaling (26). Although the most frequent cause of β -catenin deregulation in OEAs is mutation in the *CTNNB1* gene, β -catenin deregulation can also be caused by mutations in the APC, AXIN1, and AXIN2 genes. These findings imply that certain genetic alterations cause particular histological subtypes of ovarian cancer, and dysregulation of the Wnt/canonical signaling cascade might participate in development of disease (14, 27).

2.2 Epidemiology and Histopathology

Ovarian cancer is the most common gynecological cancer in North America, but accounting for only 4% of all female cancers (28). This happens as a result of ovarian cancers being missed when they are still in the early stages. The International

Federation of Gynecology and Obstetrics (FIGO) staging method is used to classify ovarian cancer, and stage 3 is where it is most frequently found. Stage 3 is linked to a 23-41% five-year survival rate (29). The OSE is estimated to be the source of 80–85% of ovarian neoplasms, which can be classified into the following histopathological subtypes: endometrioid, mucinous, and serous. Each of these histological subtypes accounts for roughly 75%, 20%, and 2% of epithelial ovarian tumors (30). The spectrum of tumors that make up each histological category of ovarian epithelial neoplasia ranges from benign to malignant. Serous and mucinous tumors often have a 70 to 85% benign propensity, whereas endometrioid and clear cell tumors typically have a high malignant propensity (31). The coelomic mesothelium, which also gives rise to the Müllerian duct and its descendants, is the source of the OSE. The uterus, the cervix, the fallopian tubes, and the top two thirds of the vagina are examples of derivatives of the Müllerian duct (32). The OSE was derived embryologically from the coelomic mesothelium, which may have given the OSE the inherent aptitude for divergent differentiation along the Müllerian pathways, which is thought to explain the considerable morphological variety of ovarian epithelial neoplasms. Interesting similarities between ovarian epithelial neoplasm subtypes and Müllerian duct derivatives have been found by histological analysis of ovarian carcinomas (33). More particular, mucinous tumors are said to mimic endocervical epithelium while serous tumors resemble fallopian tube epithelium. As the name suggests, endometrioid tumors mimic the uterine endometrium, whereas clear cell tumors are thought to resemble mesonephric cells (34). At cellular/molecular levels, ovarian tumors are extraordinarily varied as 90% or more exhibit an epithelial histology. These epithelial ovarian cells that come from the coelomic epithelium were assumed to be the source of the disease (35). Clinically, complicated cystic masses are common clinical presentation of epithelial ovarian cancer cells that usually disseminate to peritoneal, diaphragmatic, and hepatic lymph nodes (36). Unlike other malignancies, ovarian cancer cells are able to spread throughout the peritoneal cavity without encountering any anatomical barriers (37).

2.3 Causes and pathogenesis

The recurrent injury to the ovarian surface epithelium during postovulatory healing can lead to a gradual buildup of genetic anomalies. According to gonadotropin theory, stimulation of surface epithelial cells by pituitary gonadotropin surges at ovulation and their continued high concentrations after menopause leads to accumulation of genetic alterations and carcinogenesis (38). According to other theory, ovulation, surface-epithelium repair, talc or asbestos exposure, endometriosis, pelvic inflammatory disease, and measles are all increase the risk of developing epithelial ovarian cancer. This increased risk may be explained by inflammation and changes in redox potential during these processes (39, 40). Women with impaired BRCA1 and BRCA2 function have a decreased ability to repair genetic damage, which increases their risk of disease (41). Since cyclooxygenase-2 (COX2) is necessary for the inflammatory environment in which ovulation takes place, this notion supports investigating the chemopreventive potential of COX2 inhibitors (42). Luteinizing hormone preovulatory surge causes an increase in cytokine expression, which causes a differentiation in follicle cells. Invaginations and inclusion cysts are also induced during ovulation. Under an influence of ongoing cytokine and hormonal stimulation, cyst cells can develop to take on Müllerian features, proliferate, and become ciliated or secretory. These cells eventually develop genetic anomalies (43). Rete ovarii tubules at the ovary's hilus, near the transition from mesothelium to OSE, also contain ciliated and secretory cells and have the capacity to enlarge to form cysts. It is uncertain if either cyst type contains malignant cells (44). It is yet unclear how any ovarian stem or progenitor cells might contribute to the development of epithelial ovarian cancer. In one study, the authors were able to demonstrate genes that encoded for epithelial ovarian cancer (EOC) biomarkers that previously thought to be over-expressed (45). However, it is crucial to note that none of these EOC markers, perhaps with the exception of CA-125, displayed appreciable over-expression. The results had normal ovarian tissue and the other of which contained EOC as well as normal and cancerous fallopian tissue. These findings support the idea that fallopian fimbriae are the source of EOC, and they also show that markers that previously over-expressed in EOC are fallopian origin but not ovarian (46).

2.4. Diagnosis and staging

The FIGO method, as amended in 1988, is the most widely used staging system. Its discoveries are primarily the result of surgical investigation. Only 20% of ovarian cancer patients receive a diagnosis when the tumor is still localized, despite the fact that 80% of people with this disease experience symptoms (47). The lack of specificity in these symptoms is fundamental cause, and the typical symptoms include mild stomach discomfort, stool irregularities, and unexplained weight loss to severe abdominal enlargement (48). Early diagnosis is challenging due to the symptoms generality and overlaps with other more prevalent gastrointestinal, genitourinary, and gynecological diseases (49). The most widely utilized biomarkers for early

identification of disease is the cancer antigen 125 (CA125). The using of CA-125 as a biomarker or tumor marker could increase in blood of some patients with specific types of cancers or other benign conditions. High specificity can be attained through tracking CA125 levels over time or using used in conjunction with transvaginal sonography (TVS), (50, 51). Currently, the most accurate method for determining whether ovarian cancer surgery is feasible is surgical staging, and survival of the patient depends critically on eradication of all macroscopically discernible tumor symptoms, preferably completely (52).

2.5 Biomarkers

There are currently few methods for evaluating suspected instances of ovarian cancer in their early stages, which rely on pelvic ultrasonography and the CA-125 test with a sensitivity and specificity level between 70-80% (50-52). The MUC16 gene in humans produces the protein known as CA-125, commonly referred to as mucin 16 or MUC16. The mucin family of glycoproteins includes MUC16 (53). MUC16 is a part of epithelia of female reproductive tract, respiratory tract, and ocular surface. Since MUC16 is heavily glycosylated, it produces a hydrophilic environment on the apical membrane of epithelial cells that functions as a lubricating barrier against foreign particles and pathogenic pathogens. The most used biomarker for ovarian cancer screening is CA-125 (54). Human epididymis protein 4 (HE4) was found as a biomarker for ovarian cancer by Moore and associates in 2008 (55). The WAP-four disulfide core domain 2 (WFDC2) gene is responsible for encoding and producing of HE4 protein, and discovered later to be expressed in some ovarian malignancies in addition to being expressed in pulmonary epithelial cells (56). Using a combination of CA-125 levels, HE4 expression, and menopausal state, a risk of ovarian malignancy algorithm (ROMA) was created based on these findings and is now used to predict the existence of malignant ovarian cancer. HE4 and CA-125 in the ROMA, in particular, have been linked to a higher sensitivity than any single biomarker (57).

2.6. Molecular genetics

Ovarian tumors from various patients have been discovered to contain a number of genetic and epigenetic alterations. Ovarian cancer, like other solid tumors, arises from a single cell that has acquired five or more genetic mutations or more (58). For ovarian oncogenesis, neoplastic cells can avoid going through the apoptotic process when TP53 is lost or mutated, which allows them to continue dividing even after experiencing DNA damage and accumulating more genomic rearrangements (59). George et al. (2016) reported that 10-15% of ovarian cancer cases are caused by genetic flaws in the DNA repair genes themselves (60). The lifetime risk of developing ovarian cancer for bearers of these genes ranges from 30-60% for BRCA1, 15-30% for BRCA2, and 7% for MLH1 and MLH2. The efficient repair of DNA double strand breaks through homologous recombination depends on BRCA1 and BRCA2 (61). Ovarian tumors have been shown to poss about 15 distinct oncogenes, with 11 of them exhibiting genomic amplification (62). Small G-protein Rab25 is one of the most frequently increased genes in ovarian tumors. Rab25 controls apoptosis, autophagy, aggression, motility, and apoptosis. Additionally, it has been demonstrated that it mediates survival in the face of stress, such as glucose deprivation, serum depletion, UV radiation and chemotherapy (63). Aurora kinase is necessary to growing of cells. Another class of amplified oncogenes are frequently found in ovarian cancer are necessary for cytokinesis to be completed, and when they are dysregulated, they produce aneuploidy, which is a characteristic of many malignancies (64). According to studies linking genetic mutations to clinical presentation and tumor morphological characteristics, neoplasm progress is sequential manner of adenoma (65, 66). These cancers characterize by genetic alterations in KRAS, BRAF, PIK3CA, and LOH on Xq indicating that the RAF/MAPK pathway is dysregulated in development of these tumors. Also, the mutant KRAS produces cause inappropriate activation of the PI3K, RAF/MAPK, and RAL-GEF pathways (67). High-grade tumors, on other hand, start off as surface epithelium, and expand quickly and don't have any precursor lesions that may be identified morphologically. High-grade serous cancers are more likely to exhibit TP53 mutations, possible BRCA1 and BRCA2 aberrations, LOH on 7q and 9p, and BRCA1 and BRCA2 mutations (68).

2.7. Signaling pathways

Essential pathway components including PIK3CA and AKT2, as well as PTEN-inactivating mutations, can all lead to the PI3K pathway becoming activated. This pathway is also activated in several malignancies by receptor tyrosine kinase autocrine or paracrine signaling (69). Multiple pathways underlie the promotion of cell growth and survival by PI3K-Akt signaling (70). The pro-apoptotic Bcl-2 family members Bad and Bax inhibited by Akt, can prevent the transcription of anti-apoptotic genes from being negatively regulated by the transcription factor NF- κ B and pro-survival genes. Akt blocks p53 - mediated apoptosis by phosphorylating Mdm2. By inhibiting transcription factors, Akt also lessens the production of proteins that promote cell death (71). This comprises the 4E-binding protein (4E-BP) and the ribosomal protein S6 kinases (p70S6K). The latter causes the production of eIF4E, a translation initiation factor with anti-apoptotic and transforming properties *in vitro* (72). Ovarian cancer xenograft development has been demonstrated to be inhibited by PI3K and Akt, and these inhibitors may enhance cytotoxic effect of chemotherapy (73). This results in over-expression of genes that induce angiogenesis caused by JAK2 activation (74). The presence of nuclear localization of phosphorylated STAT3 in more than 70% of ovarian tumors is linked to a decline in overall survival (75). Transcriptional regulator More than half of ovarian tumors have constitutively active NF- κ B, which has been linked to cancer survival, proliferation, and metastasis. The Rel family proteins p50 and p65 form heterodimeric complexes to form the bulk of NF- κ B (76). A number of cytokines, including IL-1 and TNF, as well as growth factor eventually cause NF- κ B releasing, and in more than 50% of ovarian tumors, the over-expressed protein kinase MEKK 3 might trigger it. Additionally, it causes the over-expression of genes that crucial for controlling cell survival. In line with these findings, NF-B has been linked to cancer cells resistance to chemotherapeutic agent and radiation-induced apoptosis (76, 77).

2.8. Wnt gene

2.8.1. Signaling Molecule

On human chromosomes, there 19 Wnt genes were discovered and identified. All Wnt proteins are roughly the same size, with molecular weights between 39 and 46 kDa (78). Wnts are secreted glycoproteins with lipid modifications that are palmitoylated on a conserved cysteine to help them target the cell membrane. Associating with glycosaminoglycans in the extracellular matrix after being secreted, Wnt proteins stay near to the cell surface (79). Additionally, findings have shown that Wnts can act as long-range morphogenic signals that are concentration-dependent and can influence distant neighbors (80). However, the precise mechanism underpinning this activity is still unknown.

2.8.2. Wnt Signaling Pathways

Wnts can activate different intracellular signaling pathways when they bind to Frizzled (Fz) receptors including Wnt/ β -catenin canonical signaling system, the Wnt/ Ca^{2+} pathway, and the planar cell polarity (PCP) pathway (14). Each of these pathways can be activated by various combinations of Wnt molecules, resulting in various cellular responses (81). This discovery was made in research where Wnt/, Wnt3/, Wnt8 or Wnt8 was over-expressed in *Xenopus*, which in turn caused Wnt/-catenin pathway to be specifically stimulated. Contrarily, it was discovered that over-expression of Wnt4, Wnt5, or Wnt1 stimulated the Wnt / Ca^{2+} pathway (14, 82). In over-expression tests, Fz receptors can be divided into similar functional groups depending on their capacity to activate one pathway as opposed to another. This association between certain Wnts and the intracellular pathway anticipate to be stimulated, despite the fact that these classifications may offer a useful tool for anticipating the activity of Wnts and Fzs (83).

2.8.3. Wnt/ β -catenin pathway

The canonical Wnt/ β -catenin pathway is the subject of extensive research and is covered in detail in the preceding chapter. Briefly, cytosolic buildup of β -catenin is necessary for signaling through this route. Multi-protein destruction complex targets β -catenin for deterioration in absence of Wnt activation. As a result, the destruction complex is repelled by wnt signaling causes a buildup of β -catenin in cytosol and subsequent activation of target genes (14, 84, 85).

2.8.4. Wnt/Ca²⁺ pathway

Studies on *Xenopus* have suggested that a certain subset of Wnt ligands and Fz receptors, including Wnt4, Wnt5, Wnt/1, and Fz2, can activate the Wnt/Ca²⁺ pathway (79). A heterotrimeric G protein is activated as part of this pathway, and calcium/calmodulin-regulated kinase II (CamKII) and protein kinase C are also activated (PKC), (86). Although the downstream targets of CamKII and PKC are not yet known, it has been demonstrated that in *Xenopus*, activating the Wnt/Ca²⁺ pathway can inhibit the Wnt/ β -catenin pathway (87). Uncertainty exists over the degree at which this interaction takes place and whether or not it also occurs in mammals (14, 84).

2.8.5. Planar cell polarity pathway

Through the control of cytoskeletal architecture, the Wnt/PCP signaling pathway affects cell polarity, which can be deduced from cell shape and mobility. As it necessitates Fz receptors and Dvl molecules, the PCP pathway overlaps with the traditional signaling pathway. However, it differs from downstream processes since it does not include axin, GSK3, or β -catenin (84, 88).

2.8.6. Wnt/ β -catenin signaling and tumorigenesis

The Wnt/canonical signaling cascade's primary and crucial element is β -catenin. It is also a crucial part of the cadherin cell adhesion complex because it uses β -catenin and α -actinin to connect cadherins at the plasma membrane with the actin cytoskeleton (84). Axin, a protein that facilitates the development of the catenin degradation complex is affected by CTNNB1 gene (14). However, the loss of APC alone is insufficient for the development of carcinomas and metastasis. Additional mutations in various pathways, including as mutations that activate Ras and mutations that cause p53 to lose its function, are also necessary (89). Furthermore, β -catenin is only found in the nucleus of late dedifferentiated, mesenchyme-like tumor cells; it is not present in the plasma membrane in the center of primary tumors. As a result, additional signaling elements might be necessary for catenin to accumulate in nucleus. Additionally, phosphorylation of key tyrosine constituent parts might cause adherens junctions to dissociate (90). Interestingly, malignancies have been found to over-express tyrosine kinases and contain mutations in tyrosine phosphatase genes that may accelerate these phosphorylation processes. Increased β -catenin-dependent transcription is correlated with loss of cell adhesions, and associated with malignant prognosis (91).

2.9. Cadherins and ovarian tumorigenesis

The family of integral membrane glycoproteins known as cadherins mediates homotypic calcium-dependent cell-cell attachment. The E-, N-, and P-cadherins are three examples of the traditional cadherins that belong to this family of cell adhesion molecules (CAMs). These cadherins have been demonstrated to be crucial for the development of intercellular junctions, the maintenance of cell polarity, and the preservation of tissue integrity (92). Changes in cadherin expression or function have been suggested as crucial elements in carcinogenesis due to the fact that cell-cell interactions are frequently disordered in tumors and must be weakened in invasive and metastatic malignancies (73). Additionally, as cadherins play a role in tissue morphogenesis and cell sorting, improper or changed levels of cadherin expression within tissues may offer a mechanism for the development of metaplastic lesions or dysplastic derivatives which could be responsible for the establishment of the preneoplastic state (93).

E-cadherin gene has been classified as a tumor suppressor gene as a result of research linking E-cadherin deletion or decreased expression to the establishment of poorly-differentiated invasive carcinomas (94). Numerous studies have demonstrated that differentiating between various histological subtypes of ovarian cancer can be followed for expression of E- and N-cadherin. For example, serous and endometrioid ovarian tumors express both cadherin subtypes, in contrast to mucinous ovarian cancers, which only express E-cadherin and not N-cadherin. Interestingly, serous and mucinous subtypes of ovarian cancer trail endometrioid ovarian tumors in terms of risk for malignancy (95). It has demonstrated that EMTs take place as epithelial malignancies grow, giving these tumors an increase in motility and invasiveness. The Wnt/canonical signaling pathway among many other carcinogenic pathways induce EMT, a hall mark feature of which is the down-regulation of epithelial (E)-cadherin (96). It's interesting to note that research suggests the opposite might apply to ovarian cancer. It is believed that the OSE repeatedly rupturing and mending after ovulation causes carcinogenesis, which starts with the development of inclusion cysts in the ovarian stroma (97). These and ovarian cancers have both been demonstrated to express E-cadherin, whereas normal

OSE has only been demonstrated to express N-cadherin (98). Additionally, research looking at the transfection of regular OSE cells with E-cadherin has demonstrated that these cells start to produce inclusions cysts and take on a more invasive nature (99, 100). These findings in contrast with the well-known phenomena of EMT that takes place during cancer in other organs and suggest that the OSE may undergo a mesenchymal to epithelial phenotypic transition (79, 101, 102).

4. Conclusion

Ovarian cancer is one of the most deadly diseases impacting women worldwide. Inherently resistant cell populations have recently been connected to tumor recurrence, unsatisfactory treatment outcomes, and consequent disease relapse. However, the lack of models that faithfully reproduce the variety of ovarian cancer has hindered the study of these phenotypically and functionally complex cancer cells up to this point.

References

- [1] Jiang, Y., Li, C., Chen, L., Wang, F., and Zhou, X. (2017). Potential role of retinoids in ovarian physiology and pathogenesis of polycystic ovary syndrome. *Clinica chimica acta*, 469, 87-93.
- [2] Lambertini, M., Horicks, F., Del Mastro, L., Partridge, A. H., and Demeestere, I. (2019). Ovarian protection with gonadotropin-releasing hormone agonists during chemotherapy in cancer patients: from biological evidence to clinical application. *Cancer treatment reviews*, 72, 65-77.
- [3] Oduwole, O. O., Huhtaniemi, I. T., and Misrahi, M. (2021). The roles of luteinizing hormone, follicle-stimulating hormone and testosterone in spermatogenesis and folliculogenesis revisited. *International journal of molecular sciences*, 22(23), 12735.
- [4] Diamanti-Kandarakis, E., and Dunaif, A. (2012). Insulin resistance and the polycystic ovary syndrome revisited: an update on mechanisms and implications. *Endocrine reviews*, 33(6), 981-1030.
- [5] Parikesit, D., Mochtar, C. A., Umbas, R., and Hamid, A. R. A. H. (2016). The impact of obesity towards prostate diseases. *Prostate international*, 4(1), 1-6.
- [6] Torre, L. A., Trabert, B., DeSantis, C. E., Miller, K. D., Samimi, G., Runowicz, C. D., and Siegel, R. L. (2018). Ovarian cancer statistics, 2018. *CA: a cancer journal for clinicians*, 68(4), 284-296.
- [7] Rustin, G. J., Van Der Burg, M. E., Griffin, C. L., Guthrie, D., Lamont, A., Jayson, G. C., and Swart, A. M. (2010). Early versus delayed treatment of relapsed ovarian cancer (MRC OV05/EORTC 55955): a randomised trial. *The Lancet*, 376(9747), 1155-1163.
- [8] Bookman, M. A. (2016). Optimal primary therapy of ovarian cancer. *Annals of Oncology*, 27, i58-i62.
- [9] Marcus, C. S., Maxwell, G. L., Darcy, K. M., Hamilton, C. A., and McGuire, W. P. (2014). Current approaches and challenges in managing and monitoring treatment response in ovarian cancer. *Journal of Cancer*, 5(1), 25-30.
- [10] Brown, J. S., O'Carrigan, B., Jackson, S. P., and Yap, T. A. (2017). Targeting DNA repair in cancer: beyond PARP inhibitors. *Cancer discovery*, 7(1), 20-37.
- [11] Prat, J. (2017). Pathology of borderline and invasive cancers. *Best practice and research Clinical obstetrics and gynaecology*, 41, 15-30.
- [12] Song, H., Cicek, M. S., Dicks, E., Harrington, P., Ramus, S. J., Cunningham, J. M., and Pharoah, P. D. (2014). The contribution of deleterious germline mutations in BRCA1, BRCA2 and the mismatch repair genes to ovarian cancer in the population. *Human molecular genetics*, 23(17), 4703-4709.
- [13] Zhu, J., Zhang, S., Gu, L., and Di, W. (2012). Epigenetic silencing of DKK2 and Wnt signal pathway components in human ovarian carcinoma. *Carcinogenesis*, 33(12), 2334-2343.
- [14] Arend, R. C., Londoño-Joshi, A. I., Straughn Jr, J. M., and Buchsbaum, D. J. (2013). The Wnt/ β -catenin pathway in ovarian cancer: a review. *Gynecologic oncology*, 131(3), 772-779.
- [15] Ge, X., and Wang, X. (2010). Role of Wnt canonical pathway in hematological malignancies. *Journal of hematology and oncology*, 3, 1-6.
- [16] Basu, M., and Roy, S. S. (2013). Wnt/ β -catenin pathway is regulated by PITX2 homeodomain protein and thus contributes to the proliferation of human ovarian adenocarcinoma cell, SKOV-3. *Journal of Biological Chemistry*, 288(6), 4355-4367.
- [17] Russo, A., Czarnecki, A. A., Dean, M., Modi, D. A., Lantvit, D. D., Hardy, L., and Burdette, J. E. (2018). PTEN loss in the fallopian tube induces hyperplasia and ovarian tumor formation. *Oncogene*, 37(15), 1976-1990.
- [18] An, Q., Zhou, Y., Han, C., Zhou, Y., Li, F., and Li, D. (2017). BTG3 overexpression suppresses the proliferation and invasion in epithelial ovarian cancer cell by regulating AKT/GSK3 β / β -catenin signaling. *Reproductive Sciences*, 24, 1462-1468.

- [19] Elbadawy, M., Usui, T., Yamawaki, H., and Sasaki, K. (2019). Emerging roles of C-Myc in cancer stem cell-related signaling and resistance to cancer chemotherapy: a potential therapeutic target against colorectal cancer. *International journal of molecular sciences*, 20(9), 2340.
- [20] Vriend, J., and Nachtigal, M. W. (2021). Ubiquitin proteasome pathway transcriptome in epithelial ovarian cancer. *Cancers*, 13(11), 2659.
- [21] Usui, A., Ko, S. Y., Barengo, N., and Naora, H. (2014). P-cadherin promotes ovarian cancer dissemination through tumor cell aggregation and tumor–peritoneum interactions. *Molecular Cancer Research*, 12(4), 504-513.
- [22] Bhagat, R., Premalata, C. S., Shilpa, V., Pallavi, V. R., Ramesh, G., Vijay, C. R., and Krishnamoorthy, L. (2013). Altered expression of β -catenin, E-cadherin, and E-cadherin promoter methylation in epithelial ovarian carcinoma. *Tumor Biology*, 34, 2459-2468.
- [23] Fracalossi, A. C. C., de Souza Silva, M., Oshima, C. T. F., and Ribeiro, D. A. (2010). Wnt/ β -catenin signalling pathway following rat tongue carcinogenesis induced by 4-nitroquinoline 1-oxide. *Experimental and molecular pathology*, 88(1), 176-183.
- [24] Tonelli, F. (2020). Colon cancer: pathology and natural history. In *Hormone Replacement Therapy and Cancer*. CRC Press. Pp: 161-171.
- [25] Kossai, M., Leary, A., Scoazec, J. Y., and Genestie, C. (2018). Ovarian cancer: a heterogeneous disease. *Pathobiology*, 85(1-2), 41-49.
- [26] Acton, Q. A. (2013). *Ovaries-Advances in Research and Application: 2013 Edition: Scholarly Paper*.
- [27] Mostowska, A., Pawlik, P., Sajdak, S., Markowska, J., Pawłowska, M., Lianeri, M., and Jagodzinski, P. P. (2014). An analysis of polymorphisms within the Wnt signaling pathway in relation to ovarian cancer risk in a Polish population. *Molecular diagnosis and therapy*, 18, 85-91.
- [28] Stewart, C., Ralyea, C., and Lockwood, S. (2019, April). Ovarian cancer: an integrated review. In *Seminars in oncology nursing* (Vol. 35, No. 2, pp. 151-156). WB Saunders.
- [29] Rutten, M. J., Van De Vrie, R., Bruining, A., Spijkerboer, A. M., Mol, B. W., Kenter, G. G., and Buist, M. R. (2015). Predicting surgical outcome in patients with International Federation of Gynecology and Obstetrics stage III or IV ovarian cancer using computed tomography: a systematic review of prediction models. *International Journal of Gynecologic Cancer*, 25(3).
- [30] Wang, Y., Liu, J., Lin, B., Wang, C., Li, Q., Liu, S., and Iwamori, M. (2011). Study on the expression and clinical significances of Lewis y antigen and integrin α v, β 3 in epithelial ovarian tumors. *International journal of molecular sciences*, 12(6), 3409-3421.
- [31] Pierson, W. E., Peters, P. N., Chang, M. T., Chen, L. M., Quigley, D. A., Ashworth, A., and Chapman, J. S. (2020). An integrated molecular profile of endometrioid ovarian cancer. *Gynecologic Oncology*, 157(1), 55-61.
- [32] Auersperg, N. (2013). Ovarian surface epithelium as a source of ovarian cancers: unwarranted speculation or evidence-based hypothesis?. *Gynecologic oncology*, 130(1), 246-251.
- [33] Falcone, E. L. (2006). Ovarian cancer cells exhibit aberrations in the Wnt signaling pathway, which affect cell proliferation and cadherin expression. Master of Science Thesis, Division of Experimental Medicine, McGill University.
- [34] Stolnicu, S., Barsan, I., Hoang, L., Patel, P., Terinte, C., Pesci, A., and Soslow, R. A. (2018). International Endocervical Adenocarcinoma Criteria and Classification (IECC): a new pathogenetic classification for invasive adenocarcinomas of the endocervix. *The American journal of surgical pathology*, 42(2), 214.
- [35] Auersperg, N. (2011). The origin of ovarian carcinomas: a unifying hypothesis. *International Journal of Gynecological Pathology*, 30(1), 12-21.
- [36] Mogi, K., Yoshihara, M., Iyoshi, S., Kitami, K., Uno, K., Tano, S., and Kajiyama, H. (2021). Ovarian cancer-associated mesothelial cells: Transdifferentiation to minions of cancer and orchestrate developing peritoneal dissemination. *Cancers*, 13(6), 1352.
- [37] Plagens-Rotman, K., Połocka-Molińska, M., Merks, P., Gwoździcka-Piotrowska, M., Kędzia, W., and Jarząbek-Bielecka, G. (2021). Problems in gynaecologic oncology in girls and young women—an outline of selected issues. *Clinical and Experimental Obstetrics and Gynecology*, 48(4), 795-799.
- [38] Tomao, F., Lo Russo, G., Spinelli, G. P., Stati, V., Prete, A. A., Prinzi, N., and Tomao, S. (2014). Fertility drugs, reproductive strategies and ovarian cancer risk. *Journal of ovarian research*, 7, 1-8.
- [39] Kisielewski, R., Mazurek, A., Laudański, P., and Tołwińska, A. (2013). Inflammation and ovarian cancer—current views. *Ginekologia polska*, 84(4), 1-15.

- [40] Matulonis, U. A., Sood, A. K., Fallowfield, L., Howitt, B. E., Sehouli, J., and Karlan, B. Y. (2016). Ovarian cancer. *Nature reviews Disease primers*, 2(1), 1-22.
- [41] Tan, D. S., and Kaye, S. B. (2015). Chemotherapy for patients with BRCA1 and BRCA2-mutated ovarian cancer: Same or different?. *American Society of Clinical Oncology Educational Book*, 35(1), 114-121.
- [42] Zhang, X., Yan, K., Deng, L., Liang, J., Liang, H., Feng, D., and Ling, B. (2019). Cyclooxygenase 2 promotes proliferation and invasion in ovarian cancer cells via the PGE2/NF- κ B pathway. *Cell Transplantation*, 28(1_suppl), 1S-13S.
- [43] Crespo, D., Goetz, F. W., and Planas, J. V. (2015). Luteinizing hormone induces ovulation via tumor necrosis factor α -dependent increases in prostaglandin F2 α in a nonmammalian vertebrate. *Scientific reports*, 5(1), 1-12.
- [44] Apperson, K. D., Bird, K. E., Cherian, G., and Löhr, C. V. (2017). Histology of the ovary of the laying hen (*Gallus domesticus*). *Veterinary Sciences*, 4(4), 66.
- [45] Ning, L., Long, B., Zhang, W., Yu, M., Wang, S., Cao, D., and Lang, J. (2018). Circular RNA profiling reveals circEXOC6B and circN4BP2L2 as novel prognostic biomarkers in epithelial ovarian cancer. *International journal of oncology*, 53(6), 2637-2646.
- [46] Darelus, A., Kristjansdottir, B., Dahm-Kähler, P., and Strandell, A. (2021). Risk of epithelial ovarian cancer Type I and II after hysterectomy, salpingectomy and tubal ligation—A nationwide case-control study. *International Journal of Cancer*, 149(8), 1544-1552.
- [47] Tafe, L. J., Garg, K., Chew, I., Tornos, C., and Soslow, R. A. (2010). Endometrial and ovarian carcinomas with undifferentiated components: clinically aggressive and frequently underrecognized neoplasms. *Modern Pathology*, 23(6), 781-789.
- [48] Donovan, K. A., Donovan, H. S., Cella, D., Gaines, M. E., Penson, R. T., Plaxe, S. C., and Wenzel, L. (2014). Recommended patient-reported core set of symptoms and quality-of-life domains to measure in ovarian cancer treatment trials. *JNCI: Journal of the National Cancer Institute*, 106(7), dju128.
- [49] Lheureux, S., Braunstein, M., and Oza, A. M. (2019). Epithelial ovarian cancer: evolution of management in the era of precision medicine. *CA: a cancer journal for clinicians*, 69(4), 280-304.
- [50] Doubeni, C. A., Doubeni, A. R., and Myers, A. E. (2016). Diagnosis and management of ovarian cancer. *American family physician*, 93(11), 937-944.
- [51] Kamal, R., Hamed, S., Mansour, S., Mounir, Y., and Abdel Sallam, S. (2018). Ovarian cancer screening—ultrasound; impact on ovarian cancer mortality. *The British journal of radiology*, 91(1090), 20170571.
- [52] Elzarkaa, A. A., Shaalan, W., Elemam, D., Mansour, H., Melis, M., Malik, E., and Soliman, A. A. (2018). Peritoneal cancer index as a predictor of survival in advanced stage serous epithelial ovarian cancer: a prospective study. *Journal of gynecologic oncology*, 29(4).
- [53] Liu, J., Li, L., Luo, N., Liu, Q., Liu, L., Chen, D., and Xi, X. (2021). Inflammatory signals induce MUC16 expression in ovarian cancer cells via NF- κ B activation. *Experimental and Therapeutic Medicine*, 21(2), 1-1.
- [54] Miller, E. M., Samec, T. M., and Alexander-Bryant, A. A. (2021). Nanoparticle delivery systems to combat drug resistance in ovarian cancer. *Nanomedicine: Nanotechnology, Biology and Medicine*, 31, 102309.
- [55] Hertlein, L., Stieber, P., Kirschenhofer, A., Fürst, S., Mayr, D., Hofmann, K., and Burges, A. (2012). Human epididymis protein 4 (HE4) in benign and malignant diseases. *Clinical Chemistry and Laboratory Medicine (CCLM)*, 50(12), 2181-2188.
- [56] Chen, Y., Wang, S., Liu, T., Wu, Y., Li, J. L., and Li, M. (2016). WAP four-disulfide core domain protein 2 gene (WFDC2) is a target of estrogen in ovarian cancer cells. *Journal of ovarian research*, 9(1), 1-11.
- [57] Huy, N. V. Q., Van Khoa, V., Vinh, T. Q., Tung, N. S., Thanh, C. N., and Chuang, L. (2018). Standard and optimal cut-off values of serum ca-125, HE4 and ROMA in preoperative prediction of ovarian cancer in Vietnam. *Gynecologic oncology reports*, 25, 110-114.
- [58] Kondrashova, O., Nguyen, M., Shield-Artin, K., Tinker, A. V., Teng, N. N., Harrell, M. I., and Scott, C. L. (2017). Secondary somatic mutations restoring RAD51C and RAD51D associated with acquired resistance to the PARP inhibitor rucaparib in high-grade ovarian carcinoma. *Cancer discovery*, 7(9), 984-998.
- [59] Silwal-Pandit, L., Langerød, A., and Børresen-Dale, A. L. (2017). TP53 mutations in breast and ovarian cancer. *Cold Spring Harbor perspectives in medicine*, 7(1), a026252.
- [60] George, A., Riddell, D., Seal, S., Talukdar, S., Mahamdallie, S., Ruark, E., and Rahman, N. (2016). Implementing rapid, robust, cost-effective, patient-centred, routine genetic testing in ovarian cancer patients. *Scientific reports*, 6(1), 29506.
- [61] Sessa, C., Balmaña, J., Bober, S. L., Cardoso, M. J., Colombo, N., Curigliano, G., and Paluch-Shimon, S. (2023). Risk reduction and screening of cancer in hereditary breast-ovarian cancer syndromes: ESMO Clinical Practice Guideline☆. *Annals of Oncology*, 34(1), 33-47.
- [62] Lengyel, E., Burdette, J. E., Kenny, H. A., Matei, D., Pilrose, J., Haluska, P., and Stack, M. S. (2014). Epithelial ovarian cancer experimental models. *Oncogene*, 33(28), 3619-3633.

- [63] Lall, P., Horgan, C. P., Oda, S., Franklin, E., Sultana, A., Hanscom, S. R., and Khan, A. R. (2013). Structural and functional analysis of FIP2 binding to the endosome-localised Rab25 GTPase. *Biochimica et Biophysica Acta (BBA)-Proteins and Proteomics*, 1834(12), 2679-2690.
- [64] Pérez-Fidalgo, J. A., Gambardella, V., Pineda, B., Burgues, O., Piñero, O., and Cervantes, A. (2020). Aurora kinases in ovarian cancer. *ESMO open*, 5(5), e000718.
- [65] Sun, Y., Yokoi, K., Li, H., Gao, J., Hu, L., Liu, B., and Zhang, W. (2011). NGAL Expression Is Elevated in Both Colorectal Adenoma–Carcinoma Sequence and Cancer Progression and Enhances Tumorigenesis in Xenograft Mouse Models NGAL in Tumorigenesis and Progression of Colorectal Cancer. *Clinical Cancer Research*, 17(13), 4331-4340.
- [66] Choi, J. W., Liu, H., Shin, D. H., Yu, G. I., Hwang, J. S., Kim, E. S., and Yun, J. W. (2013). Proteomic and cytokine plasma biomarkers for predicting progression from colorectal adenoma to carcinoma in human patients. *Proteomics*, 13(15), 2361-2374.
- [67] Weidle, U. H., Birzele, F., Kollmorgen, G., and Rueger, R. (2016). Mechanisms and targets involved in dissemination of ovarian cancer. *Cancer genomics and proteomics*, 13(6), 407-423.
- [68] Jung, Y. Y., Woo, H. Y., and Kim, H. S. (2019). Targeted genomic sequencing reveals novel TP53 in-frame deletion mutations leading to p53 overexpression in high-grade serous tubo-ovarian carcinoma. *Anticancer Research*, 39(6), 2883-2889.
- [69] Owusu-Brackett, N., Shariati, M., and Meric-Bernstam, F. (2019). Role of pi3k/akt/mtor in cancer signaling. *Predictive biomarkers in oncology: applications in precision medicine*, 263-270.
- [70] Ediriweera, M. K., Tennekoon, K. H., and Samarakoon, S. R. (2019, December). Role of the PI3K/AKT/mTOR signaling pathway in ovarian cancer: Biological and therapeutic significance. In *Seminars in cancer biology* (Vol. 59, pp. 147-160). Academic Press.
- [71] Linnerth-Petrik, N. M., Santry, L. A., Moorehead, R., Jücker, M., Wootton, S. K., and Petrik, J. (2016). Akt isoform specific effects in ovarian cancer progression. *Oncotarget*, 7(46), 74820.
- [72] Taha, A. A., Koshiyama, M., Matsumura, N., Abiko, K., Yamaguchi, K., Hamanishi, J., and Mandai, M. (2018). The effect of the type of dietary protein on the development of ovarian cancer. *Oncotarget*, 9(35), 23987.
- [73] Lengyel, E. (2010). Ovarian cancer development and metastasis. *The American journal of pathology*, 177(3), 1053-1064.
- [74] Duan, Z., Foster, R., Bell, D. A., Mahoney, J., Wolak, K., Vaidya, A., and Seiden, M. V. (2006). Signal transducers and activators of transcription 3 pathway activation in drug-resistant ovarian cancer. *Clinical Cancer Research*, 12(17), 5055-5063.
- [75] Abubaker, K., Luwor, R. B., Zhu, H., McNally, O., Quinn, M. A., Burns, C. J., and Ahmed, N. (2014). Inhibition of the JAK2/STAT3 pathway in ovarian cancer results in the loss of cancer stem cell-like characteristics and a reduced tumor burden. *BMC cancer*, 14(1), 1-22.
- [76] Annunziata, C. M., Stavnes, H. T., Kleinberg, L., Berner, A., Hernandez, L. F., Birrer, M. J., and Kohn, E. C. (2010). Nuclear factor κB transcription factors are coexpressed and convey a poor outcome in ovarian cancer. *Cancer*, 116(13), 3276-3284.
- [77] Cvrljevic, A. N., Butt, U., Huhtinen, K., Grönroos, T. J., Böckelman, C., Lassus, H., and Westermarck, J. (2022). Ovarian Cancers with Low CIP2A Tumor Expression Constitute an APR-246–Sensitive Disease Subtype. *Molecular cancer therapeutics*, 21(7), 1236-1245.
- [78] Nguyen, V. H. L., Hough, R., Bernaudo, S., and Peng, C. (2019). Wnt/β-catenin signalling in ovarian cancer: insights into its hyperactivation and function in tumorigenesis. *Journal of ovarian research*, 12, 1-17.
- [79] Teeuwssen, M., and Fodde, R. (2019). Wnt signaling in ovarian cancer stemness, EMT, and therapy resistance. *Journal of clinical medicine*, 8(10), 1658.
- [80] Waghmare, I., and Page-McCaw, A. (2018). Wnt signaling in stem cell maintenance and differentiation in the *Drosophila* Germarium. *Genes*, 9(3), 127.
- [81] Van Jaarsveld, M. T. M., Helleman, J., Boersma, A. W. M., Van Kuijk, P. F., Van Ijcken, W. F., Despiere, E., and Wiemer, E. A. C. (2013). miR-141 regulates KEAP1 and modulates cisplatin sensitivity in ovarian cancer cells. *Oncogene*, 32(36), 4284-4293.
- [82] Chau, W. K., Ip, C. K., Mak, A. S. C., Lai, H. C., and Wong, A. S. T. (2013). c-Kit mediates chemoresistance and tumor-initiating capacity of ovarian cancer cells through activation of Wnt/β-catenin–ATP-binding cassette G2 signaling. *Oncogene*, 32(22), 2767-2781.

- [83] Lu, Y., Beeghly-Fadiel, A., Wu, L., Guo, X., Li, B., Schildkraut, J. M., and Long, J. (2018). A transcriptome-wide association study among 97,898 women to identify candidate susceptibility genes for epithelial ovarian cancer risk. *Cancer research*, 78(18), 5419-5430.
- [84] Arend, R. C., Londoño-Joshi, A. I., Samant, R. S., Li, Y., Conner, M., Hidalgo, B., and Buchsbaum, D. J. (2014). Inhibition of Wnt/ β -catenin pathway by niclosamide: A therapeutic target for ovarian cancer. *Gynecologic oncology*, 134(1), 112-120.
- [85] Kildal, W., Risberg, B., Abeler, V. M., Kristensen, G. B., Sudbø, J., Nesland, J. M., and Danielsen, H. E. (2005). β -catenin expression, DNA ploidy and clinicopathological features in ovarian cancer: A study in 253 patients. *European journal of cancer*, 41(8), 1127-1134.
- [86] Arang, N., and Gutkind, J. S. (2020). G Protein-Coupled receptors and heterotrimeric G proteins as cancer drivers. *FEBS letters*, 594(24), 4201-4232.
- [87] McCoy, F., Darbandi, R., Chen, S. I., Eckard, L., Dodd, K., Jones, K., and Nutt, L. K. (2013). Metabolic regulation of CaMKII protein and caspases in *Xenopus laevis* egg extracts. *Journal of Biological Chemistry*, 288(13), 8838-8848.
- [88] Zhang, K., Song, H., Yang, P., Dai, X., Li, Y., Wang, L., and Zhang, T. (2015). Silencing dishevelled-1 sensitizes paclitaxel-resistant human ovarian cancer cells via AKT/GSK-3 β / β -catenin signalling. *Cell proliferation*, 48(2), 249-258.
- [89] Rechsteiner, M., Zimmermann, A. K., Wild, P. J., Caduff, R., von Teichman, A., Fink, D., and Noske, A. (2013). TP53 mutations are common in all subtypes of epithelial ovarian cancer and occur concomitantly with KRAS mutations in the mucinous type. *Experimental and molecular pathology*, 95(2), 235-241.
- [90] Ghilardi, C., Moreira-Barbosa, C., Brunelli, L., Ostano, P., Panini, N., Lupi, M., and Giavazzi, R. (2022). PGC1 α / β expression predicts therapeutic response to oxidative phosphorylation inhibition in ovarian cancer. *Cancer Research*, 82(7), 1423-1434.
- [91] Vouri, M., and Hafizi, S. (2017). TAM receptor tyrosine kinases in cancer drug resistance. *Cancer Research*, 77(11), 2775-2778.
- [92] Quattrocchi, L., Green, A. R., Martin, S., Durrant, L., and Deen, S. (2011). The cadherin switch in ovarian high-grade serous carcinoma is associated with disease progression. *Virchows Archiv*, 459, 21-29.
- [93] Delman, D. M., Fabian, C. J., Kimler, B. F., Yeh, H., and Petroff, B. K. (2015). Effects of flaxseed lignan secoisolariciresinol diglucoside on preneoplastic biomarkers of cancer progression in a model of simultaneous breast and ovarian cancer development. *Nutrition and cancer*, 67(5), 857-864.
- [94] Wu, Q., Guo, R., Lin, M., Zhou, B., and Wang, Y. (2011). MicroRNA-200a inhibits CD133/1+ ovarian cancer stem cells migration and invasion by targeting E-cadherin repressor ZEB2. *Gynecologic oncology*, 122(1), 149-154.
- [95] Frumovitz, M., Schmeler, K. M., Malpica, A., Sood, A. K., and Gershenson, D. M. (2010). Unmasking the complexities of mucinous ovarian carcinoma. *Gynecologic oncology*, 117(3), 491-496.
- [96] Klymenko, Y., Kim, O., and Stack, M. S. (2017). Complex determinants of epithelial: mesenchymal phenotypic plasticity in ovarian cancer. *Cancers*, 9(8), 104.
- [97] Otsuka, I. (2021). Mechanisms of high-grade serous carcinogenesis in the fallopian tube and ovary: current hypotheses, etiologic factors, and molecular alterations. *International Journal of Molecular Sciences*, 22(9), 4409.
- [98] Jannesari-Ladani, F., Hossein, G., Monhasery, N., Shahoei, S. H., and Mood, N. I. (2014). Wnt5a influences viability, migration, adhesion, colony formation, E-and N-cadherin expression of human ovarian cancer cell line SKOV-3. *Folia Biologica*, 60(2), 57.
- [99] Wu, C., Cipollone, J., Maines-Bandiera, S., Tan, C., Karsan, A., Auersperg, N., and Roskelley, C. D. (2008). The morphogenic function of E-cadherin-mediated adherens junctions in epithelial ovarian carcinoma formation and progression. *Differentiation*, 76(2), 193-205.
- [100] Choi, P. W., Yang, J., Ng, S. K., Feltmate, C., Muto, M. G., Hasselblatt, K., and Ng, S. W. (2016). Loss of E-cadherin disrupts ovarian epithelial inclusion cyst formation and collective cell movement in ovarian cancer cells. *Oncotarget*, 7(4), 4110.
- [101] Davidson, B., Tropé, C. G., and Reich, R. (2012). Epithelial–mesenchymal transition in ovarian carcinoma. *Frontiers in oncology*, 2, 33.
- [102] Suster, N. K., and Virant-Klun, I. (2019). Presence and role of stem cells in ovarian cancer. *World journal of stem cells*, 11(7), 383.